



Autoimmune Gastritis: Focus on Endoscopic and Morphological Characteristics

Andrey P. Kiryukhin^{1*}, Alexander S. Tertychnyy¹, Pavel V. Pavlov¹,
Alexandr A. Fedorenko¹, Darina P. Nagornaya¹, Natalia S. Marenich²,
Ekaterina A. Losik¹, Elena Yu. Yuryeva¹, Tatiana L. Lapina¹

¹ I.M. Sechenov First Moscow State Medical University (Sechenov University), Moscow, Russian Federation

² Morozov Children's City Clinical Hospital of the Moscow Health Department, Moscow, Russian Federation

Aim: to systematize the literature data on endoscopic semiotics and morphological changes in the gastric mucosa in autoimmune gastritis.

Key points. Autoimmune gastritis is associated with an increased risk of developing adenocarcinoma and neuroendocrine tumours of the stomach. Clarification of diagnostic criteria for autoimmune gastritis is essential for gastroenterological practice. The diagnosis is based on the results of endoscopic and histological examination, and on data from laboratory tests. Isolated atrophy of the mucous membrane of the body of the stomach, the presence of difficult-to-wash creamy mucus, changes in the mucous membrane like "shed skin", and the presence of whitish globule-like foci are typical endoscopic signs of autoimmune gastritis. Widespread pseudopyloric metaplasia, focal intestinal and pseudopancreatic metaplasia, hyperplasia of the ridges of the mucous membrane of the body of the stomach and their relationship to the glandular layer as in the antrum allow during a morphological study considering clinical data to suspect and verify autoimmune gastritis.

Conclusion. During instrumental examination, knowledge of endoscopic symptoms and pathognomonic morphological changes is important for the timely diagnosis of autoimmune gastritis.

Keywords: autoimmune gastritis, endoscopy, biopsy, atrophy, *Helicobacter pylori*

Conflict of interest: the authors declare that there is no conflict of interest.

For citation: Kiryukhin A.P., Tertychny A.S., Pavlov P.V., Fedorenko A.A., Nagornaya D.P., Marenich N.S., Losik E.A., Yuryeva E.Yu., Lapina T.L. Autoimmune Gastritis: Focus on Endoscopic and Morphological Characteristics. Russian Journal of Gastroenterology, Hepatology, Coloproctology. 2024;34(1):58–69. <https://doi.org/10.22416/1382-4376-2024-34-1-58-69>

Аутоиммунный гастрит: в фокусе эндоскопические и морфологические характеристики

А.П. Кирюхин^{1*}, А.С. Тertychnyy¹, П.В. Павлов¹, А.А. Федоренко¹, Д.П. Нагорная¹, Н.С. Маренич²,
Е.А. Лосик¹, Е.Ю. Юрьева¹, Т.Л. Лапина¹

¹ ФГАОУ ВО «Первый Московский государственный медицинский университет им. И.М. Сеченова» Министерства здравоохранения Российской Федерации (Сеченовский Университет), Москва, Российская Федерация

² ГБУЗ города Москвы «Морозовская детская городская клиническая больница Департамента здравоохранения города Москвы», Москва, Российская Федерация

Цель: систематизировать литературные данные об эндоскопической семиотике и морфологических изменениях слизистой оболочки желудка при аутоиммунном гастрите.

Основные положения. Аутоиммунный гастрит ассоциирован с повышенным риском развития аденокарциномы и нейроэндокринных опухолей желудка. Уточнение диагностических критериев аутоиммунного гастрита имеет существенное значение для гастроэнтерологической практики. Установление диагноза основано на результатах эндоскопического и гистологического исследования, на данных лабораторных тестов. Изолированная атрофия слизистой оболочки тела желудка, наличие трудноотмываемой кремоподобной слизи, изменение слизистой оболочки по типу «сброшенной кожи», наличие белесых глобулоподобных очагов служат типичными эндоскопическими признаками аутоиммунного гастрита. Распространенная псевдопилорическая метаплазия, очаговая кишечная и псевдопанкреатическая метаплазия, гиперплазия валиков слизистой оболочки тела желудка и их соотношение к железистому слою как в антральном отделе позволяют при морфологическом исследовании с учетом клинических данных заподозрить и верифицировать аутоиммунный гастрит.

Закключение. При инструментальном обследовании знание эндоскопических симптомов и патогномоничных морфологических изменений является важным для своевременного диагноза аутоиммунного гастрита.

Ключевые слова: аутоиммунный гастрит, эндоскопия, биопсия, атрофия, *Helicobacter pylori*

Конфликт интересов: авторы заявляют об отсутствии конфликта интересов.

Для цитирования: Кирюхин А.П., Тертычный А.С., Павлов П.В., Федоренко А.А., Нагорная Д.П., Маренич Н.С., Лосик Е.А., Юрьева Е.Ю., Лапина Т.Л. Аутоиммунный гастрит: в фокусе эндоскопические и морфологические характеристики. Российский журнал гастроэнтерологии, гепатологии, колопроктологии. 2024;34(1):58–69. <https://doi.org/10.22416/1382-4376-2024-34-1-58-69>

Atrophic gastritis is a precancerous condition, and its timely diagnosis is crucial for cancer prevention [1–3]. The frequency of *H. pylori*-associated gastritis is decreasing globally, which some researchers attribute to improved hygiene levels and positive outcomes of eradication therapy with antibacterial agents [4, 5]. Considering this trend, the study of autoimmune gastritis (AIG) as an independent significant cause of gastric mucosal atrophy becomes increasingly relevant [6, 7].

AIG is an organ-specific immune-mediated disease characterized by the destruction of gastric parietal cells, leading to the loss of intrinsic factor (Castle factor) and a reduction in gastric acid production. These changes result in iron and vitamin B₁₂ malabsorption [8, 9]. The detection of autoantibodies against H⁺, K⁺-ATPase and intrinsic factor, reduced serum pepsinogen I, and the ratio of pepsinogen I to pepsinogen II, along with increased gastrin-17, are crucial for AIG diagnosis [4, 9–12]. AIG often coexists with other autoimmune diseases such as autoimmune thyroiditis, less frequently with type 1 diabetes, Addison's disease, vitiligo, and chronic urticaria [9, 11].

The role of *H. pylori* infection in the initiation and progression of the autoimmune process in the stomach is still unclear [8]. For instance, there is speculation that *H. pylori*, through antigenic mimicry, may trigger autoimmune gastritis [13]. Antibodies to parietal cells can be detected in *H. pylori*-positive individuals without AIG [8]. According to one clinical study, up to 62 % of AIG patients had a positive *H. pylori* test result [14].

In a study by E.A. Losik, a high frequency of autoimmune gastritis was found in patients with autoimmune thyroiditis (51.9 %). It was noted that the prevalence of AIG increases with age (27.9 % in patients aged 20–39 years and 53.8 % in patients aged 70–79 years among those surveyed). The frequency of *H. pylori* infection in AIG patients did not differ from population rates, although individuals with pronounced atrophic changes in the gastric body encountered the infectious agent significantly less often [15]. Hematological abnormalities were identified in 25.4 % of patients with high titres of antibodies against parietal cells. Cases of anemia

(11.3 %) manifested as both macrocytic and microcytic types.

An increase in the average volume of red blood cells and a low level of vitamin B₁₂ were found in 16.9 % of patients, while a decrease in hemoglobin levels was observed in 25 % of cases [10].

Patients with AIG have an increased risk of developing gastric adenocarcinoma and type 1 neuroendocrine tumours of the stomach. During endoscopic observation of patients with pernicious anemia (from 0 to 20 years), the frequency of gastric adenocarcinoma was 3 %, and type 1 neuroendocrine tumours were 4 % [16]. In a cohort study involving 275 patients with atrophic gastritis of the gastric body observed for an average of 5 years (ranging from 1 to 17 years), the annual incidence of gastric cancer/high-grade dysplasia was 0.5 %, low-grade dysplasia — 0.6 %, and type 1 neuroendocrine tumour — 2.8 %. Age over 60 (risk ratio — 4.7), intestinal metaplasia in the absence of pseudopyloric metaplasia (risk ratio — 4.3), and pernicious anemia (risk ratio — 4.3) were associated with a high risk of gastric cancer/high-grade dysplasia and low-grade dysplasia [6]. The potential of AIG as a precancerous condition requires further investigation.

P. Kriş et al. did not find a higher prevalence of antibodies against parietal cells and intrinsic factor in patients diagnosed with gastric adenocarcinoma compared to the control group [17]. During long-term follow-up of AIG patients without a history of *H. pylori* infection, no increased risk of gastric cancer was observed compared to the control group, and the authors suggest that this risk may be associated with preceding or unrecognized ongoing *H. pylori* infection [18].

Endoscopic semiotics of autoimmune gastritis

Japanese researchers conducted a retrospective analysis of endoscopic signs of autoimmune gastritis (AIG) in 222 patients and identified key features of AIG, including: predominant localization of mucosal atrophy in the gastric body area (frequency — 96 %); clear visualization of submucosal vessels and flattening/disappearance of longitudinal folds; variable absence of intestinal metaplasia in the mucosa of the antral region [19–21].

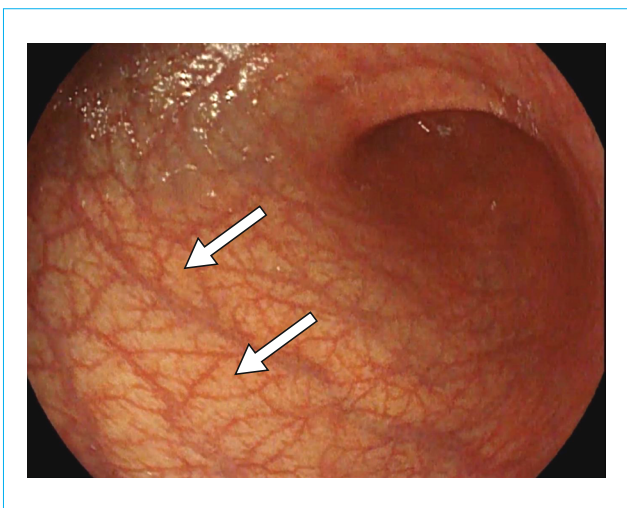


Figure 1. Endophotography during video esophagogastroduodenoscopy in white light: the mucous membrane of the gastric body is diffusely thinned with visible submucosal vessels of various sizes (arrows). (Hereinafter, endophotographies are presented from the archives of the Department of Diagnostic and Treatment Endoscopy of the University Clinical Hospital No. 2 of Sechenov University)

Рисунок 1. Эндофотография при видеоэзофагогастроуденоскопии в белом свете: слизистая оболочка тела желудка диффузно истончена, с видимыми подслизистыми сосудами различного калибра (стрелки). (Здесь и далее представлены эндофотографии из архива отделения диагностической и лечебной эндоскопии Университетской клинической больницы № 2 Семеновского университета.)

Figure 1 and 2 show endoscopic images of a patient suffered from AIG with atrophy predominantly confined to the gastric body.

When evaluating the data from video esophagogastroduodenoscopy and identifying signs of gastric mucosal atrophy, its stage is determined based on the Kimura – Takemoto classification or the modified EGA (Endoscopic Grading of Atrophy) scale [22–24]. This classification and scale were recommended by the Kyoto Consensus for assessing the prevalence of mucosal atrophy in *H. pylori*-associated gastritis. However, in the case of AIG, considering the absence or minimal atrophic changes in the antral part of the stomach, the accuracy of these scales is limited, according to some researchers [25].

Additionally, five types of changes in the gastric body mucosa, characterized by the nature of visualized remnants of glandular epithelium during endoscopic examination, are described: flat-raised localized (frequency – 48.6 %); pseudopolypoid – raised islands of remaining non-thinned mucosa with pronounced atrophic phenomena at

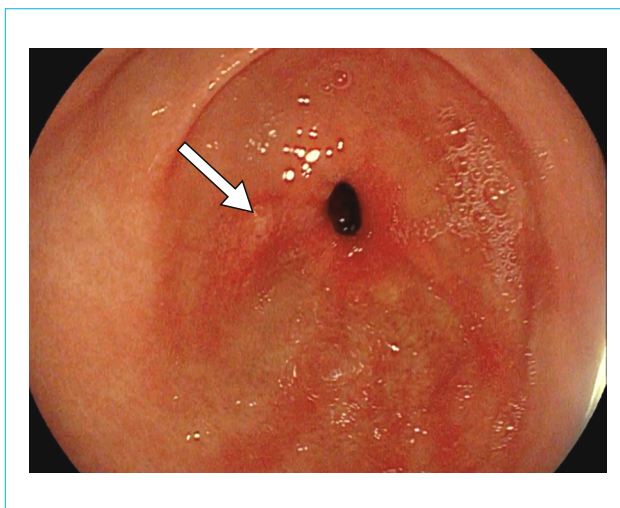


Figure 2. Endophotography during video esophagogastroduodenoscopy in white light (the same patient as in Fig. 1): the mucous membrane of the antrum of the stomach with linear hyperemia, a single epithelizing defect of the mucous membrane under fibrin (arrow)

Рисунок 2. Эндофотография при видеоэзофагогастроуденоскопии в белом свете (тот же пациент, что и на рис. 1): слизистая оболочка антрального отдела желудка с линейной гиперемией, единичным эпителизирующимся дефектом слизистой оболочки под фибрином (стрелка)

their periphery (22.9 %); island-like (18.6 %); extensively focal (7.1 %); granular (2.9 %) [19, 20].

Figures 3 and 4 show endophotographs demonstrating foci of preserved mucous membrane against the background of atrophy in a patient with AIG.

The mucous membrane of the antral part of the stomach in AIG may be intact or show nonspecific changes such as focal hyperemia, raised erosions, and, in some patients, foci of circular wrinkling of the mucous membrane (in 22 % of AIG cases). Against the background of hypochlorhydria and bacterial contamination, the presence of viscous, difficult-to-wash cream-like mucus with a pale-yellow colour can be visualized (frequency – 32.4 %) [19, 20]. This feature is presented in Figure 5.

When evaluating the pit pattern of the mucous membrane in a narrow spectrum, the phenomenon of the disappearance of centrally located gland orifices with changes in the mucous membrane resembling “shed skin” (frequency – 59 %) is observed [20, 26]. An additional endoscopic marker for the presence of AIG may be the identification of white globule-like foci during routine video endoscopic examination (“white globe appearance”), described as white spherical structures with a diameter of up to 1 mm with microvessels above them, reflecting the location of these structures beneath the gastric epithelium

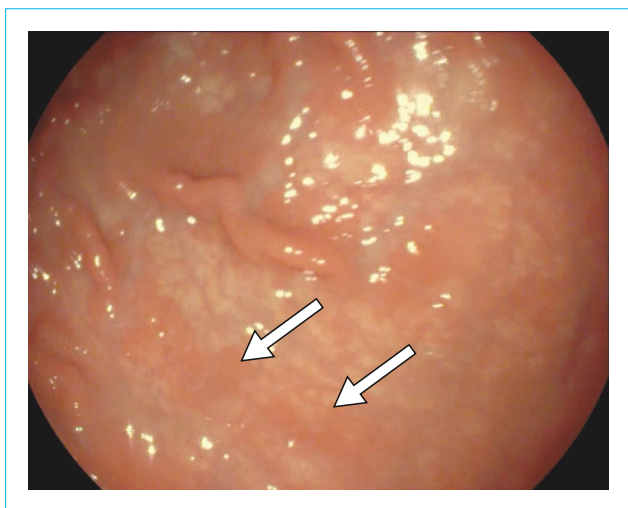


Figure 3. Endophotography during video esophagogastroduodenoscopy in white light: diffusely thinned mucous membrane of the body of the stomach is whitish in colour with islands of intact mucous membrane (arrows)

Рисунок 3. Эндофотография при видеоэзофагогастродуоденоскопии в белом свете: диффузно истонченная слизистая оболочка тела желудка белесого цвета с островками интактной слизистой оболочки (стрелки)

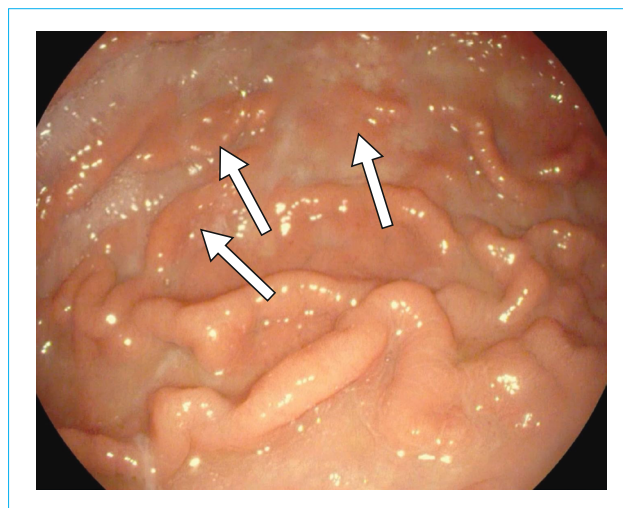


Figure 4. Endophotography during video esophagogastroduodenoscopy in white light: thinned mucous membrane of the body of the stomach is whitish in colour with raised islands of non-thinned mucous membrane of the longitudinal folds of the greater curvature of the body of the stomach (arrows)

Рисунок 4. Эндофотография при видеоэзофагогастродуоденоскопии в белом свете: истонченная слизистая оболочка тела желудка белесого цвета с приподнятыми островками неистонченной слизистой оболочки продольных складок большой кривизны тела желудка (стрелки)

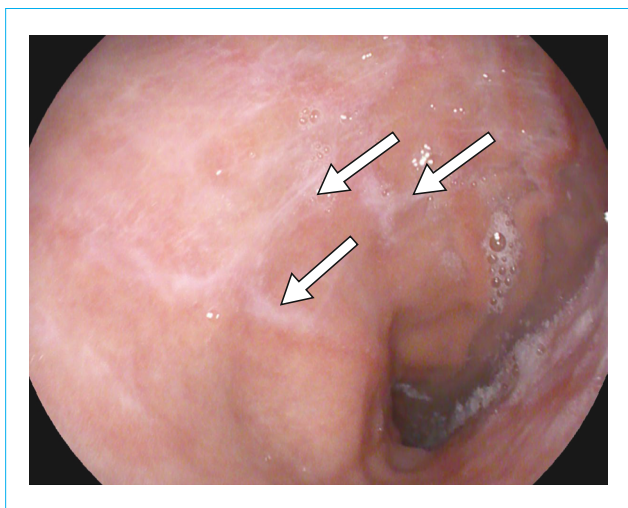


Figure 5. Endophotography during video esophagogastroduodenoscopy in white light: thinned mucous membrane of the body of the stomach with viscous, difficult-to-wash, creamy mucus of a whitish-yellow colour (arrows)

Рисунок 5. Эндофотография при видеоэзофагогастродуоденоскопии в белом свете: истонченная слизистая оболочка тела желудка с вязкой, трудноотмываемой кремоподобной слизью белесо-желтого цвета (стрелки)



Figure 6. Endophotography during video esophagogastroduodenoscopy in white light: thinned mucous membrane of the gastric body with a whitish globule-like lesion (arrow)

Рисунок 6. Эндофотография при видеоэзофагогастродуоденоскопии в белом свете: истонченная слизистая оболочка тела желудка с белесым глобулоподобным очагом (стрелка)

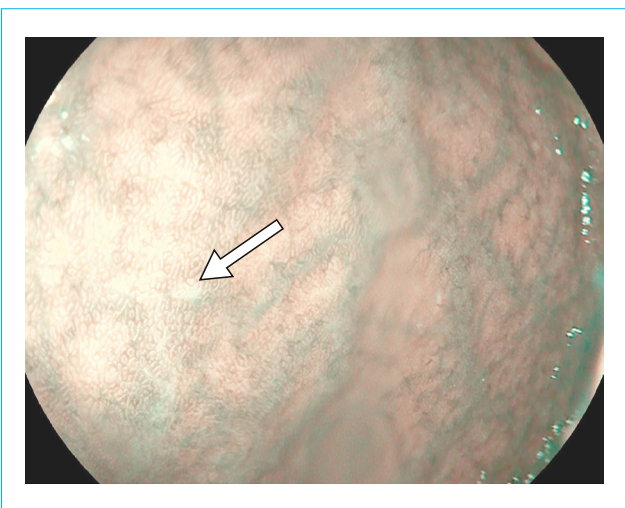


Figure 7. Endophotography during video esophagogastroduodenoscopy in a narrow spectrum of illumination: thinned mucous membrane of the body of the stomach with a whitish globule-like lesion against the background of contouring vessels (arrow)

Рисунок 7. Эндофотография при видеоэзофагогастродуоденоскопии в узком спектре освещения: истонченная слизистая оболочка тела желудка с белесым глобулоподобным очагом на фоне контурирующихся сосудов (стрелка)

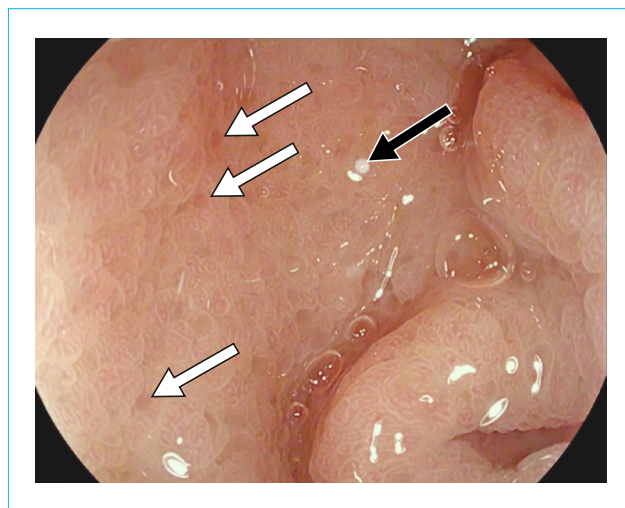


Figure 8. Endophotography during video esophagogastroduodenoscopy in white light: the mucous membrane of the body of the stomach with a whitish globule-like lesion (indicated by the black arrow) and multiple “glomus-like” neoplasms (indicated by the white arrows)

Рисунок 8. Эндофотография при видеоэзофагогастродуоденоскопии в белом свете: слизистая оболочка тела желудка с белесым глобулоподобным очагом (указана черной стрелкой) и множественными «глобус-подобными» новообразованиями (указаны белыми стрелками)

[27]. Figures 6 and 7 show endophotographs of patients with AIG with of white globule-like foci.

These formations are most commonly identified in malignant gastric lesions, but they can also be visualized in AIG, where they have an intermittent nature (may disappear within 12 months of observation) [27]. The appearance of these foci in AIG is explained by the obstruction of glandular ducts due to their destruction in the autoimmune process, leading to the formation of intramucosal cysts [27].

R. Kuvaev et al. described an additional feature of AIG during magnifying endoscopy, characterized by the presence of multiple round pale, but not white, “glomus-like” lesions (GLLs) on the mucous membrane of the gastric body, with branching capillaries on their summits (Fig. 8).

This phenomenon was observed in patients with AIG in 77 % of cases (sensitivity – 0.77; specificity – 1.00) and in 100 % of cases of AIG associated with neuroendocrine neoplasms. Morphological studies have shown that the basis of white “glomus-like” lesions lies in the hyperplasia of enterochromaffin-like cells against the background of atrophic changes [28].

Endoscopic semiotics of early manifestations of AIG has not been fully developed [29]. Literature describes individual cases of endoscopic semiotics of early AIG manifestations in the form of atrophic changes in the mucous membrane, predominantly

localized in the area of the lesser curvature of the gastric body, with the presence of reddish nodules resembling pseudopolyps, against the background of atrophic but unchanged mucous membrane along the greater curvature. In the absence of atrophic changes in the early form of AIG, edema and mosaic-like appearance of the mucous membrane of the vault and/or body of the stomach can be observed [30–32].

Patients who have undergone *H. pylori* eradication with formed atrophic changes in the mucous membrane of the antral section pose a particular challenge for the endoscopic diagnosis of AIG. In such situations, the most accurate decision is to perform a gastric mucosa biopsy according to the Sydney protocol and serological blood tests to detect antibodies to parietal cells and intrinsic factor [33].

Histological picture of autoimmune gastritis

For morphological verification of AIG during video esophagogastroduodenoscopy, biopsy of the gastric mucosa is performed according to the modified Sydney protocol, with evaluation according to the OLGA (Operative Link for Gastritis Assessment) scale: two fragments – from the antral section at a distance of 3 cm from the pylorus along the greater and lesser curvature; two fragments – from the body of the stomach in the middle of the greater curvature and at 4 cm proximal to the angle along the lesser curvature; one fragment – from the angle of the

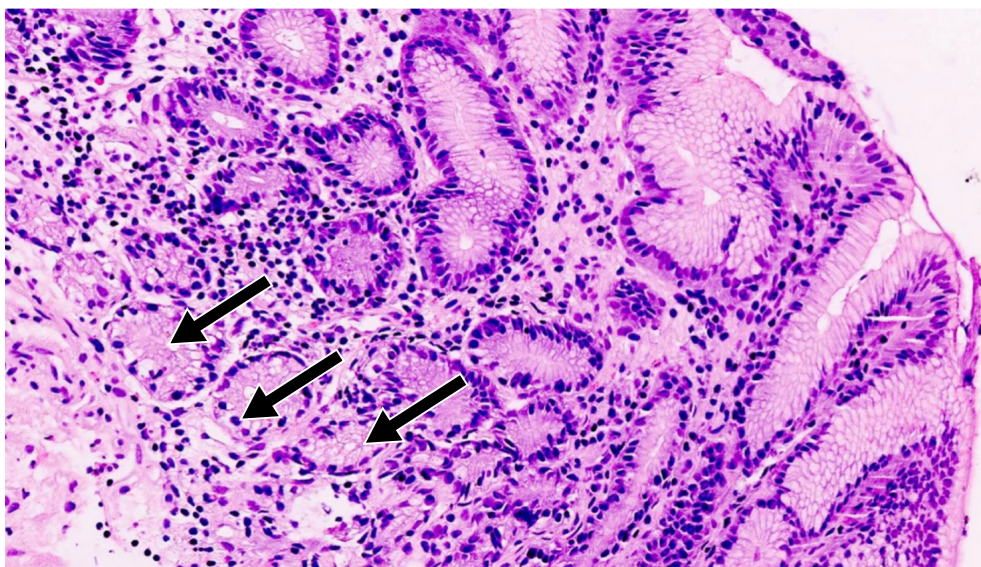


Figure 9. Chronic mild inactive atrophic gastritis of the body of the stomach with widespread pseudopyloric metaplasia (indicated by arrows) and hyperplasia of the foveolar layer. Magnification $\times 200$, stained with hematoxylin and eosin. (Microphotographs in this figure and below are from the archives of the Institute of Clinical Morphology and Digital Pathology of Sechenov University)

Рисунок 9. Хронический слабовыраженный неактивный атрофический гастрит тела желудка с распространенной псевдопилорической метаплазией (указаны стрелками) и гиперплазией фовеолярного слоя. Увеличение $\times 200$, окраска гематоксилином и эозином. (Микрофотографии на этом рисунке и далее — из архива Института клинической морфологии и цифровой патологии Сеченовского Университета)

stomach. This biopsy can be combined with material obtained from the antral section of the stomach [4].

In AIG, the inflammatory infiltration consists predominantly of mononuclear cells and is directed against the oxyntic glands with parietal cells. In the case of *H. pylori* infection, inflammatory infiltration is predominantly localized in the superficial layers of the mucous membrane, altering the architecture of the pits. In AIG, the structure of the pits can often remain intact or less damaged, but recognizable by its round or oval forms. Characteristic morphological changes in the mucous membrane in AIG include widespread pseudopyloric metaplasia, focal intestinal metaplasia, and focal pseudopancreatic metaplasia. Morphological examination of biopsies taken from the body of the stomach is practically indistinguishable from biopsies of the mucous membrane of the antral section of the stomach [7, 8]. The typical picture of chronic mild non-active atrophic gastritis of the gastric body with widespread pseudopyloric metaplasia and hyperplasia of the foveolar layer, characteristic of autoimmune gastritis, is presented in Figure 9.

The morphological characteristics of biopsies of the mucous membrane of the antral section (Fig. 10) and the body of the stomach indicate the presence of groups of mucous glands (pyloric in the antral section and pseudopyloric in the body of the stomach) and

a mild lymphoplasmacytic infiltrate. In the body of the stomach, there is hypertrophy of the rugae, and the ratio of rugae to the glandular layer is 2:1 (normal range — 1:3–4), approaching that of the antral section. This fact may be misinterpreted as a variant of normalcy and the absence of atrophy if the pathologist is not informed about the biopsy site or is unfamiliar with the morphological diagnostic features of autoimmune gastritis.

Other types of metaplasia (intestinal and pseudopancreatic) have a focal character and, in most cases, do not exceed 30 % of the biopsy area. Widespread pseudopyloric metaplasia is presented in Figure 11.

In some biopsy specimens, preserved areas of the main glands of the stomach body can be observed (Fig. 12), which, against the background of overall atrophy, may manifest as polypoid protrusions of the mucous membrane.

Polyps in the stomach body of patients with autoimmune gastritis (AIG) can also be formed by the hyperplastic foveolar layer (Fig. 13) and appear as elevated areas of rugal folds against the background of atrophic mucous membrane. They may be classified as hyperplastic polyps.

The surface pit epithelium, lining the rugae and pits, is tall and mature cylindrical. Occasionally, pits may expand, forming large cysts filled with

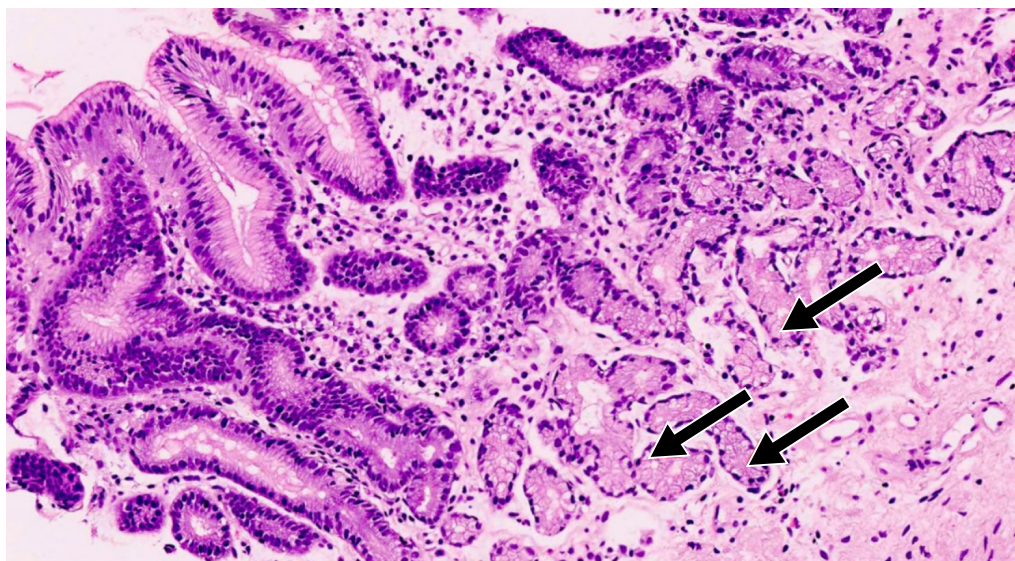


Figure 10. Chronic mild inactive superficial gastritis of the antrum of the stomach (mucous glands are indicated by arrows). Magnification $\times 200$, stained with hematoxylin and eosin

Рисунок 10. Хронический слабовыраженный неактивный поверхностный гастрит антрального отдела желудка (слизистые железы указаны стрелками). Увеличение $\times 200$, окраска гематоксилином и эозином

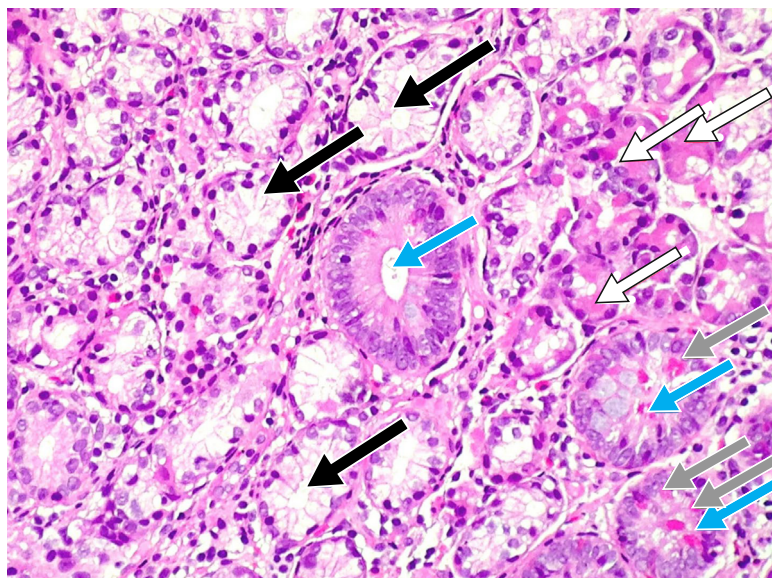


Figure 11. Widespread pseudopyloric metaplasia (black arrows) of the mucous membrane of the gastric body with individual intestinal crypts (blue arrows) with the presence of Paneth cells (grey arrows) (foci of complete intestinal metaplasia) and areas with the presence of eosinophilic staining of the cytoplasm of the glandular epithelium, resembling pancreatic acinar cells (white arrows) (focus of pseudopancreatic metaplasia). Magnification $\times 250$, stained with hematoxylin and eosin

Рисунок 11. Распространенная псевдопилорическая метаплазия (черные стрелки) слизистой оболочки тела желудка с отдельными кишечными криптами (голубые стрелки) с наличием клеток Панета (серые стрелки) (очаги полной кишечной метаплазии) и участками с наличием эозинофильного прокрашивания цитоплазмы эпителия желез, напоминающих ацинарные клетки поджелудочной железы (белые стрелки) (очаг псевдопанкреатической метаплазии). Увеличение $\times 250$, окраска гематоксилином и эозином

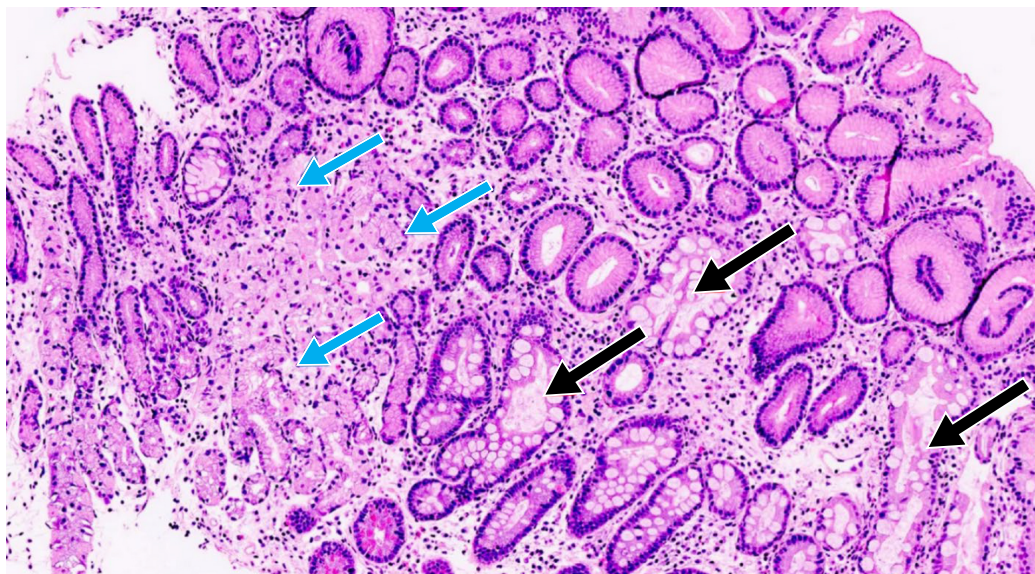


Figure 12. The preserved section of the main glands of the body of the stomach (blue arrows), along the rest of the course — mucous membrane with foci of complete intestinal metaplasia (black arrows) and hyperplasia of the foveolar layer. Magnification $\times 200$, stained with hematoxylin and eosin

Рисунок 12. Сохранившийся участок главных желез тела желудка (голубые стрелки), на остальном протяжении — слизистая оболочка с очагами полной кишечной метаплазии (черные стрелки) и гиперплазией фовеолярного слоя. Увеличение $\times 200$, окраска гематоксилином и эозином

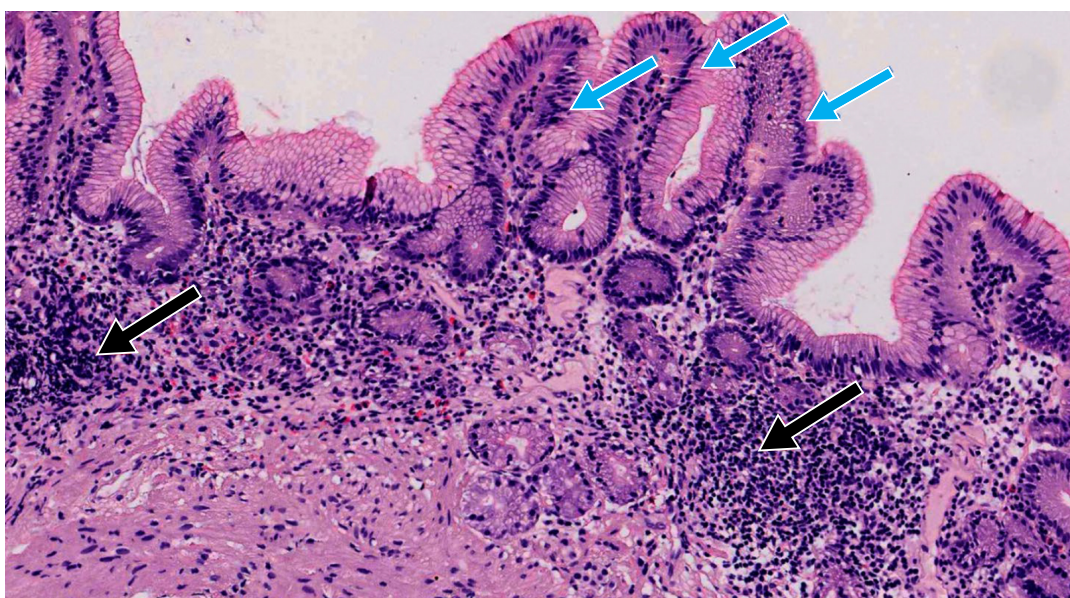


Figure 13. Areas of hyperplastic foveolar layer (blue arrows) against the background of pronounced loss of glands and widespread lymphoplasmacytic infiltration with the formation of lymphoid accumulations (black arrows). Magnification $\times 200$, stained with hematoxylin and eosin

Рисунок 13. Участки гиперплазированного фовеолярного слоя (голубые стрелки) на фоне выраженной убыли желез и распространенной лимфоплазматической инфильтрации с формированием лимфоидных скоплений (черные стрелки). Увеличение $\times 200$, окраска гематоксилином и эозином

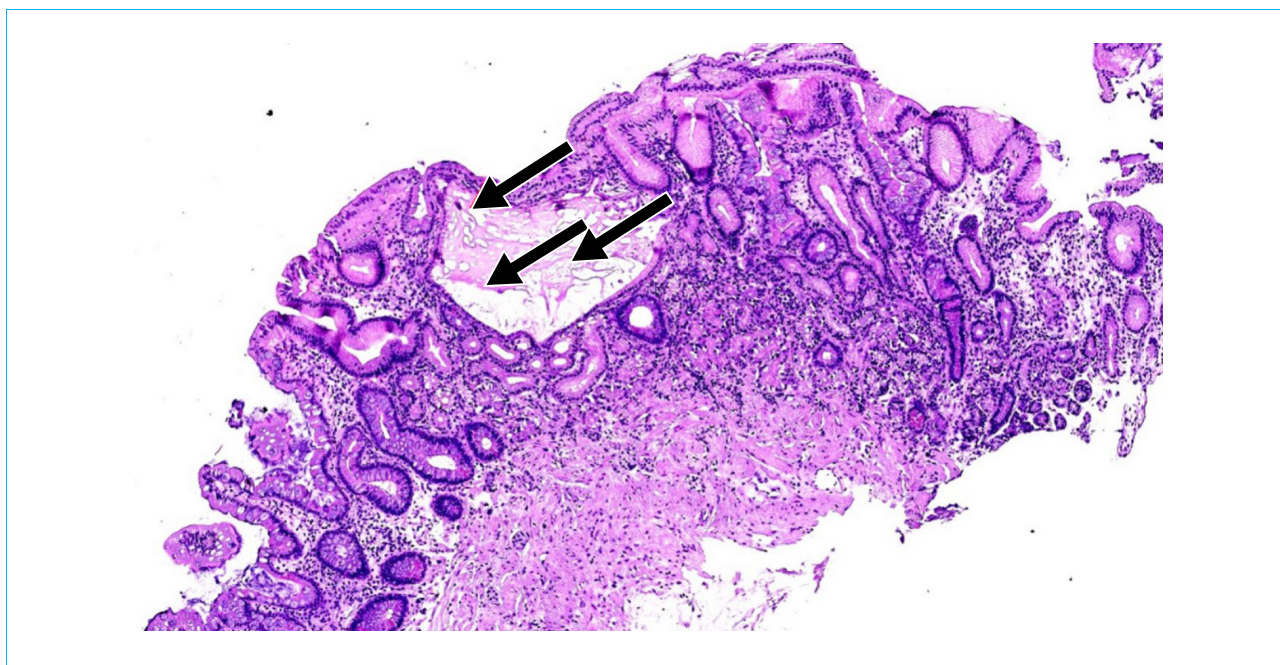


Figure 14. Chronic moderately severe inactive atrophic gastritis of the body of the stomach with focal pseudopyloric and focal complete intestinal metaplasia, hyperplasia of the foveal layer and the presence of a large cyst in the pit area with eosinophilic masses in the lumen (arrows). Magnification $\times 100$, stained with hematoxylin and eosin

Рисунок 14. Хронический умеренно выраженный неактивный атрофический гастрит тела желудка с очаговой псевдопилорической и очаговой полной кишечной метаплазией, гиперплазией фовеолярного слоя и наличием крупной кисты в зоне ямок с эозинофильными массами в просвете (стрелки). Увеличение $\times 100$, окраска гематоксилином и эозином

mucus in the form of eosinophilic masses in the lumen (Fig. 14).

Early morphological signs of AIG have a focal and mild nature, easily detectable in the morphological examination of biopsies. Pseudo-hypertrophic changes in parietal cells and small lymphoid infiltrates in the gastric body mucosa have been described upon detection of antibodies to parietal cells. With disease progression, destructive processes in the chief gland epithelium may occur, characterized by the presence of apoptotic bodies and features of emperipolesis, resulting from the immunological attack of activated T lymphocytes [8].

Conclusion

Endoscopic signs of AIG include isolated atrophy of the gastric body mucosa, the presence of viscous, difficult-to-remove cream-like mucus, changes in the mucosa resembling “shed skin”, and the presence of white globule-like lesions. Modern capabilities of endoscopic equipment and knowledge of the endoscopic AIG semiotics allow a matching

of endoscopic and histological signs. Inflammation in AIG is limited to the body and fundus of the stomach, without affecting the antrum, which distinguishes this form of gastritis from other options that lead to atrophy, for example, *H. pylori*-associated gastritis. This topography of atrophic gastritis during histological and endoscopic examination is the most important sign of the autoimmune nature of inflammation directed against the parietal cells of the oxyntic glands. Histologically, the characteristic features include widespread pseudopyloric metaplasia, focal intestinal metaplasia, focal pseudopancreatic metaplasia, hyperplasia of the rugae in the gastric body mucosa, and their ratio to the glandular layer, approaching that of the antral region. Foveal hyperplasia may endoscopically correspond to pseudopolyps. Intramucosal cysts serve as the morphological substrate of globule-like lesions, hyperplasia of enterochromaffin-like cells - “glomus-like” lesions. Early detection and verification of AIG are essential for determining patient prognosis and the subsequent management plan. It is EGD with histological examination that determines a timely diagnosis.

References / Литература

- Uemura N., Okamoto S., Yamamoto S., Matsumura N., Yamaguchi S., Yamakido M., et al. Helicobacter pylori infection and the development of gastric cancer. *N Engl J Med*. 2001;345(11):784–9. DOI: 10.1056/NEJMoa001999
- Spence A.D., Cardwell C.R., McMenamin U.C., Hicks B.M., Johnston B.T., Murray L.J., et al. Adenocarcinoma risk in gastric atrophy and intestinal metaplasia: A systematic review. *BMC Gastroenterol*. 2017;17(1):157. DOI: 10.1186/s12876-017-0708-4
- Akbari M., Tabrizi R., Kardeh S., Lankarani K.B. Gastric cancer in patients with gastric atrophy and intestinal metaplasia: A systematic review and meta-analysis. *PLoS One*. 2019;14(7):e0219865. DOI: 10.1371/journal.pone.0219865
- Ивашкин В.Т., Липина Т.Л., Маев И.В., Драпкина О.М., Козлов Р.С., Шептулин А.А. и др. Клинические рекомендации Российской гастроэнтерологической ассоциации, Научного сообщества по содействию клиническому изучению микробиома человека, Российского общества профилактики неинфекционных заболеваний, Межрегиональной ассоциации по клинической микробиологии и антимикробной химиотерапии по диагностике и лечению *H. pylori* у взрослых. *Российский журнал гастроэнтерологии, гепатологии, колопроктологии*. 2022;32(6):72–93. [Ivashkin V.T., Lipina T.L., Maev I.V., Drapkina O.M., Kozlov R.S., Sheptulin A.A., et al. Clinical Practice Guidelines of Russian Gastroenterological Association, Scientific Society for the Clinical Study of Human Microbiome, Russian Society for the Prevention of Non-Communicable Diseases, Interregional association for clinical microbiology and antimicrobial chemotherapy for *H. pylori* Diagnostics and Treatment in Adults. *Russian Journal of Gastroenterology, Hepatology, Coloproctology*. 2022;32(6):72–93. (In Russ.)]. DOI: 10.22416/1382-4376-2022-32-6-72-93
- Ивашкин В.Т., Ульянов А.И., Маев И.В., Козлов Р.С., Ливзан М.А., Абдулхаков С.Р. и др. Современные подходы к проведению эрадикационной терапии *H. pylori* у взрослых (обзор литературы и резолюция Экспертного совета). *Российский журнал гастроэнтерологии, гепатологии, колопроктологии*. 2022;32(6):7–19. [Ivashkin V.T., Ulyanin A.I., Maev I.V., Kozlov R.S., Livzan M.A., Abdulkhakov S.R., et al. Modern approaches to *H. pylori* eradication therapy in adults (literature review and resolution of Experts Council). *Russian Journal of Gastroenterology, Hepatology, Coloproctology*. 2022;32(6):7–19. (In Russ.)]. DOI: 10.22416/1382-4376-2022-32-6-7-19
- Dilaghi E., Dottori L., Pivetta G., Dalla Bella M., Esposito G., Ligato I., et al. Incidence and predictors of gastric neoplastic lesions in corpus-restricted atrophic gastritis: A single-center cohort study. *Am J Gastroenterol*. 2023;118(12):2157–65. DOI: 10.14309/ajg.0000000000002327
- Тертычный А.С., Пачуашвили Н.В., Нагорная Д.П., Липина Т.Л., Лосик Е.А., Пономарев А.Б. и др. Клинико-морфологические особенности дисплазии и раннего рака желудка, возникшего на фоне аутоиммунного гастрита. *Российский журнал гастроэнтерологии, гепатологии, колопроктологии*. 2023;33(3):16–33. [Tertychnyy A.S., Pachushvili N.V., Nagornaya D.P., Lipina T.L., Losik E.A., Ponomarev A.B., et al. Clinical and morphological features of dysplasia and early gastric cancer in the patients with autoimmune gastritis. *Russian Journal of Gastroenterology, Hepatology, Coloproctology*. 2023;33(3):16–33. (In Russ.)]. DOI: 10.22416/1382-4376-2023-33-3-16-33
- Lenti M.V., Rugge M., Lahner E., Miceli E., Toh B.H., Genta R.M., et al. Autoimmune gastritis. *Nat Rev Dis Primers*. 2020;6(1):56. DOI: 10.1038/s41572-020-0187-8
- Rustgi S.D., Bijlani P., Shah S.C. Autoimmune gastritis, with or without pernicious anemia: Epidemiology, risk factors, and clinical management. *Therap Adv Gastroenterol*. 2021;14:17562848211038771. DOI: 10.1177/17562848211038771
- Лосик Е.А., Ивашкин В.Т. Гематологические проявления аутоиммунного гастрита. *Российский журнал гастроэнтерологии, гепатологии, колопроктологии*. 2016;26(1):37–43. [Losik Ye.A., Ivashkin V.T. Hematological manifestations of autoimmune gastritis. *Russian Journal of Gastroenterology, Hepatology, Coloproctology*. 2016;26(1):37–43. (In Russ.)]. DOI: 10.22416/1382-4376-2016-26-1-37-43
- Kulnigg-Dabsch S. Autoimmune gastritis. *Wien Med Wochenschr*. 2016;166(13–14):424–30. DOI: 10.1007/s10354-016-0515-5
- Krike P., Shums Z., Poljaka I., Kikuste I., Vanags A., Tolmanis I., et al. The diagnostic value of anti-parietal cell and intrinsic factor antibodies, pepsinogens, and gastrin-17 in corpus-restricted atrophic gastritis. *Diagnostics (Basel)*. 2022;12(11):2784. DOI: 10.3390/diagnostics12112784
- Negrini R., Savio A., Poesi C., Appelmelk B.J., Bufoli F., Paterlini A., et al. Antigenic mimicry between *Helicobacter pylori* and gastric mucosa in the pathogenesis of body atrophic gastritis. *Gastroenterology*. 1996;111(3):655–65. DOI: 10.1053/gast.1996.v111.pm8780570
- Annibale B., Negrini R., Caruana P., Lahner E., Grossi C., Bordini C., et al. Two-thirds of atrophic body gastritis patients have evidence of *Helicobacter pylori* infection. *Helicobacter*. 2001;6(3):225–33. DOI: 10.1046/j.1083-4389.2001.00032.x
- Лосик Е.А. Клинико-морфологическая характеристика аутоиммунного гастрита: дисс. канд. мед. наук. М.: ФГАОУ ВО Первый Московский государственный медицинский университет им. И.М. Сеченова Минздрава России (Сеченовский Университет), 2018. [Losik E.A. Clinical and morphological characteristics of autoimmune gastritis: dissertation thesis of Cand. Sci. (Med). Moscow: I.M. Sechenov First Moscow State Medical University (Sechenov University), 2018. (In Russ.)].
- Sjöblom S.M., Sipponen P., Miettinen M., Karonen S.L., Järvinen H.J. Gastroscopic screening for gastric carcinoids and carcinoma in pernicious anemia. *Endoscopy*. 1988;20(2):52–6. DOI: 10.1055/s-2007-1018130
- Krike P., Appel M.S., Shums Z., Poljaka I., Kojalo I., Rudzite D., et al. Autoimmune gastritis serological biomarkers in gastric cancer patients. *Eur J Cancer Prev*. 2024;33(1):29–36. DOI: 10.1097/CEJ.0000000000000826
- Rugge M., Bricca L., Guzzinati S., Sacchi D., Pizzi M., Savarino E., et al. Autoimmune gastritis: Long-term natural history in naïve *Helicobacter pylori*-negative patients. *Gut*. 2023;72(1):30–8. DOI: 10.1136/gutjnl-2022-327827
- Terao S., Suzuki S., Yaita H., Kurahara K., Shunto J., Furuta T., et al. Multicenter study of autoimmune gastritis in Japan: Clinical and endoscopic characteristics. *Dig Endosc*. 2020;32(3):364–72. DOI: 10.1111/den.13500
- Kamada T., Maruyama Y., Monobe Y., Haruma K. Endoscopic features and clinical importance of autoimmune gastritis. *Dig Endosc*. 2022;34(4):700–13. DOI: 10.1111/den.14175
- Kato M., Uedo N., Toth E., Shichijo S., Maekawa A., Kanesaka T., et al. Differences in image-enhanced endoscopic findings between *Helicobacter pylori*-associated and autoimmune gastritis. *Endosc Int Open*. 2021;9:E22–30. DOI: 10.1055/a-1287-9767
- Kimura K., Takemoto T. An endoscopic recognition of the atrophic border and its significance in chronic gastritis. *Endoscopy*. 1969;1(3):87–97. DOI: 10.1055/s-0028-1098086
- Miwata T., Quach D.T., Hiyama T., Aoki R., Le H.M., Tran P.L., et al. Interobserver and intraobserver agreement for gastric mucosa atrophy. *BMC Gastroenterol*. 2015;15:95. DOI: 10.1186/s12876-015-0327-x
- Song J.H., Kim S.G., Jin E.H., Lim J.H., Yang S.Y. Risk factors for gastric tumorigenesis in underlying gastric mucosal atrophy. *Gut Liver*. 2017;11(5):612–9. DOI: 10.5009/gnl16488
- Annibale B., Esposito G., Lahner E. A current clinical overview of atrophic gastritis. *Expert Rev Gastroenterol*

- Hepatol.* 2020;14(2):93–102. DOI: 10.1080/17474124.2020.1718491
26. Maruyama Y., Yoshii M., Kageoka M. Features of magnifying endoscopic findings in type A gastritis. *Stomach Intest.* 2018;53:1516–21.
 27. Iwamuro M., Tanaka T., Kanzaki H., Kawano S., Kawahara Y., Okada H. Two cases of white globe appearance in autoimmune atrophic gastritis. *Case Rep Gastrointest Med.* 2018;2018:7091520. DOI: 10.1155/2018/7091520
 28. Kuvaev R., Kashin S., Kraynova E. A new indicator for autoimmune gastritis by magnifying image enhanced endoscopy: “Glomus like” lesions of the gastric corpus. *Dig Endosc.* 2022;34(Suppl 1):52.
 29. Hirata Y. Endoscopy opens the door to a new era of autoimmune gastritis research. *Dig Endosc.* 2020;32(3):323–5. DOI: 10.1111/den.13539
 30. Kotera T., Oe K., Kushima R., Haruma K. Multiple pseu-dopolyps presenting as reddish nodules are a characteristic endoscopic finding in patients with early-stage autoimmune gastritis. *Intern Med.* 2020;59:2995–3000. DOI: 10.2169/internalmedicine.4637-20
 31. Ayaki M., Aoki R., Mastunaga T., Manabe N., Fujita M., Kamada T., et al. Endoscopic and upper gastrointestinal barium X-ray radiography images of early-stage autoimmune gastritis: A report of two cases. *Intern Med.* 2021;60(11):1691–6. DOI: 10.2169/internalmedicine.6328-20
 32. Kishino M., Yao K., Hashimoto H., Nitta H., Kure R., Yamamoto A., et al. A case of early autoimmune gastritis with characteristic endoscopic findings. *Clin J Gastroenterol.* 2021;14(3):718–24. DOI: 10.1007/s12328-021-01351-4
 33. Nishizawa T., Yoshida S., Watanabe H., Toyoshima A., Kataoka Y., Takahashi Y., et al. Clue of diagnosis for autoimmune gastritis. *Digestion.* 2021;102(6):903–10. DOI: 10.1159/000516624

Information about the authors

Andrey P. Kiryukhin* — Cand. Sci. (Med.), Endoscopist, Department of Diagnostic and Therapeutic Endoscopy, University Clinical Hospital No. 2, I.M. Sechenov First Moscow State Medical University (Sechenov University).
Contact information: a.p.kiryukhin@gmail.com;
119435, Moscow, Pogodinskaya str., 1, build. 1.
ORCID: <https://orcid.org/0000-0001-5685-8784>

Alexander S. Tertychnyy — Dr. Sci. (Med.), Professor, Head of the Laboratory of Electron Microscopy and Immunohistochemistry, Institute of Clinical Morphology and Digital Pathology, I.M. Sechenov First Moscow State Medical University (Sechenov University).
Contact information: atertychnyy@yandex.ru;
119435, Moscow, Pogodinskaya str., 1, build. 1.
ORCID: <https://orcid.org/0000-0001-5635-6100>

Pavel V. Pavlov — Cand. Sci. (Med.), Head of the Department of Diagnostic and Therapeutic Endoscopy, University Clinical Hospital No. 2, I.M. Sechenov First Moscow State Medical University (Sechenov University).
Contact information: pvpavlov@yandex.ru;
119435, Moscow, Pogodinskaya str., 1, build. 1.
ORCID: <https://orcid.org/0000-0002-4391-5441>

Alexandr A. Fedorenko — Endoscopist, Department of Diagnostic and Therapeutic Endoscopy, University Clinical Hospital No. 2, I.M. Sechenov First Moscow State Medical University (Sechenov University).
Contact information: shyra10@mail.ru;
119435, Moscow, Pogodinskaya str., 1, build. 1.
ORCID: <https://orcid.org/0000-0002-1792-5638>

Сведения об авторах

Кирюхин Андрей Павлович* — кандидат медицинских наук, врач-эндоскопист отделения диагностической и лечебной эндоскопии Университетской клинической больницы № 2 ФГАОУ ВО «Первый Московский государственный медицинский университет им. И.М. Сеченова» Министерства здравоохранения Российской Федерации (Сеченовский Университет).
Контактная информация: a.p.kiryukhin@gmail.com;
119435, г. Москва, ул. Погодинская, 1, стр. 1.
ORCID: <https://orcid.org/0000-0001-5685-8784>

Тертычный Александр Семенович — доктор медицинских наук, профессор, заведующий лабораторией электронной микроскопии и иммуногистохимии Института клинической морфологии и цифровой патологии ФГАОУ ВО «Первый Московский государственный медицинский университет им. И.М. Сеченова» Министерства здравоохранения Российской Федерации (Сеченовский Университет).
Контактная информация: atertychnyy@yandex.ru;
119435, г. Москва, Абрикосовский пер., 1, стр. 1.
ORCID: <https://orcid.org/0000-0001-5635-6100>

Павлов Павел Владимирович — кандидат медицинских наук, заведующий отделением диагностической и лечебной эндоскопии Университетской клинической больницы № 2 ФГАОУ ВО «Первый Московский государственный медицинский университет им. И.М. Сеченова» Министерства здравоохранения Российской Федерации (Сеченовский университет).
Контактная информация: pvpavlov@yandex.ru;
119435, г. Москва, ул. Погодинская, 1, стр. 1
ORCID: <https://orcid.org/0000-0002-4391-5441>

Федоренко Александр Алексеевич — врач-эндоскопист отделения диагностической и лечебной эндоскопии Университетской клинической больницы № 2 ФГАОУ ВО «Первый Московский государственный медицинский университет им. И.М. Сеченова» Министерства здравоохранения Российской Федерации (Сеченовский Университет).
Контактная информация: shyra10@mail.ru;
119435, г. Москва, ул. Погодинская, 1, стр. 1.
ORCID: <https://orcid.org/0000-0002-1792-5638>

* Corresponding author / Автор, ответственный за переписку

Darina P. Nagornaya — Postgraduate, Institute of Clinical Morphology and Digital Pathology, I.M. Sechenov First Moscow State Medical University (Sechenov University).
Contact information: ndp.msk@ya.ru;
119435, Moscow, Abrikosovskiy lane, 1, build. 1.
ORCID: <https://orcid.org/0000-0002-5907-6195>

Natalia S. Marenich — Endoscopist of the Endoscopy Department, Morozov Children's City Clinical Hospital of the Moscow Health Department.
Contact information: nataliamarenich@mail.ru;
119049, Moscow, 4th Dobryninskiy lane, 1/9.
ORCID: <https://orcid.org/0000-0002-4391-9776>

Ekaterina A. Losik — Cand. Sci. (Med.), Cardiologist, Clinic of Propaedeutics of Internal Diseases, Gastroenterology and Hepatology named after V.Kh. Vasilenko, I.M. Sechenov First Moscow State Medical University (Sechenov University).
Contact information: kollezion@yandex.ru;
119435, Moscow, Pogodinskaya str., 1, build. 1.
ORCID: <https://orcid.org/0000-0002-3528-7983>

Elena Yu. Yuryeva — Head of the Department of Gastroenterology, Clinic of Propaedeutics of Internal Diseases, Gastroenterology and Hepatology named after V.Kh. Vasilenko, I.M. Sechenov First Moscow State Medical University (Sechenov University).
Contact information: doctorlena97@yandex.ru;
119435, Moscow, Pogodinskaya str., 1, build. 1.
ORCID: <https://orcid.org/0000-0002-0131-3063>

Tatiana L. Lapina — Cand. Sci. (Med.), Associate Professor at the Department of Propaedeutics of Internal Diseases, I.M. Sechenov First Moscow State Medical University (Sechenov University).
Contact information: tatlapina@gmail.com;
119991, Moscow, Pogodinskaya str., 1, build. 1.
ORCID: <https://orcid.org/0000-0003-4456-8725>

Нагорная Дарина Павловна — аспирант Института клинической морфологии и цифровой патологии ФГАОУ ВО «Первый Московский государственный медицинский университет им. И.М. Сеченова» Министерства здравоохранения Российской Федерации (Сеченовский Университет).
Контактная информация: ndp.msk@ya.ru;
119435, г. Москва, Абрикосовский пер., 1, стр. 1.
ORCID: <https://orcid.org/0000-0002-5907-6195>

Маренич Наталья Сергеевна — врач-эндоскопист эндоскопического отделения ГБУЗ города Москвы «Морозовская детская городская клиническая больница Департамента здравоохранения города Москвы».
Контактная информация: nataliamarenich@mail.ru;
119049, г. Москва, 4-й Добрынинский пер., 1/9.
ORCID: <https://orcid.org/0000-0002-4391-9776>

Лосик Екатерина Александровна — кандидат медицинских наук, врач-кардиолог Клиники пропедевтики внутренних болезней, гастроэнтерологии и гепатологии им. В.Х. Василенко ФГАОУ ВО «Первый Московский государственный медицинский университет им. И.М. Сеченова» Министерства здравоохранения Российской Федерации (Сеченовский Университет).
Контактная информация: kollezion@yandex.ru;
119435, г. Москва, ул. Погодинская, 1, стр. 1.
ORCID: <https://orcid.org/0000-0002-3528-7983>

Юрьева Елена Юрьевна — заведующая отделением гастроэнтерологии Клиники пропедевтики внутренних болезней, гастроэнтерологии и гепатологии им. В.Х. Василенко ФГАОУ ВО «Первый Московский государственный медицинский университет им. И.М. Сеченова» Министерства здравоохранения Российской Федерации (Сеченовский университет).
Контактная информация: doctorlena97@yandex.ru;
119435, г. Москва, ул. Погодинская, 1, стр. 1.
ORCID: <https://orcid.org/0000-0002-0131-3063>

Лапина Татьяна Львовна — кандидат медицинских наук, доцент кафедры пропедевтики внутренних болезней, гастроэнтерологии и гепатологии ФГАОУ ВО «Первый Московский государственный медицинский университет им. И.М. Сеченова» Министерства здравоохранения Российской Федерации (Сеченовский Университет).
Контактная информация: tatlapina@gmail.com;
119435, г. Москва, ул. Погодинская, 1, стр. 1.
ORCID: <https://orcid.org/0000-0003-4456-8725>

Submitted: 09.04.2023 Accepted: 09.09.2023 Published: 29.02.2024
Поступила: 09.04.2023 Принята: 09.09.2023 Опубликовано: 29.02.2024